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AUG 07 2006


PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: John G. Babish, *et al.*
Application No.: 10/789,814
Filing Date: February 27, 2004
Docket Number: 068911-0075
Title: SYNERGISTIC ANTI-INFLAMMATORY
PHARMACEUTICAL COMPOSITIONS AND METHODS
OF USE
Examiner: Shobha Kantamneni
Art Unit: 1617

CERTIFICATE OF TRANSMISSION

I hereby certify that this correspondence is being deposited with the United States Postal Service as First Class Mail in an envelope addressed to MAIL STOP AMENDMENT, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450 on the date indicated below.

Date: 08/07/06
Angelo J. Mignanelli
ERIN M. OLSONMAIL STOP AMENDMENT

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450
Sir:

DECLARATION PURSUANT TO 37 C.F.R. § 1.131

I, John G. Babish declare as follows:

1) I am Dr. John G. Babish, Executive Vice President of Metaproteomics, LLC.
I have held this position since August 2002.

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2) I have Doctorate and Masters degrees, respectively, in Biochemistry and Chemistry from Cornell University, as well as a Bachelor degree in Biochemistry from The Pennsylvania State University. A copy of my Curriculum Vitae is attached as Exhibit A.

3) I am also an inventor named in several domestic and foreign patent applications including U.S. Application Nos. 10/141,085; 10/789,814; 10/789,817; 10/988,393; 10/480,145; 10/484,123; 10/881,404; 10/774,048; 10/464,834; 10/234,002 and 09/952,632 and issued foreign and domestic patents, including U.S. Patent Nos. 6,140,063; 5,506,420; 6,629,835; 6,733,793 and 6,908,630.

4) On the basis of 30 years of training and experience, I am an expert in the art of molecular biology, more specifically, that aspect of molecular biology involving signal transduction. I was a faculty member at the College of Veterinary Medicine, Cornell University for 17 years. As Professor of Pharmacology and Toxicology, my research program involved the elucidation of mechanisms by which xenobiotics affect signaling pathways in normal and transformed cells. Using the tools of molecular biology such as monoclonal antibodies, northern and western blotting and enzyme-linked immunoassays, my research program developed cell-based assays for the identification of small molecules directed at inhibiting selected cellular functions. Findings from these studies were used to identify potential anti-viral and anti-neoplastic pharmacophores from natural products. My research has also identified both positive and negative drug-drug and drug-nutrient interactions.


5) I understand that in the course of the February 7, 2006 Office Action during prosecution in the above-captioned application, Examiner Shobha Kantamneni rejected claims 1 – 7 under 35 U.S.C. § 103(a), as allegedly being unpatentable over Kuhrts (US 2004/0137096, PTO-892) for reasons of obviousness. I respectfully submit that the instant invention was conceived prior to the January 9, 2003 filing date of the cited Kuhrts application and diligently researched until the February 27, 2004.

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6) The basis to assert that the instant invention was invented prior to the Kuhrt reference cited is supported by copies of laboratory notebook pages dated from June 2002 through December 2003 showing research on the synergistic, anti-inflammatory effects of reduced isoalpa acids (RIAA) and isoalpa acids (IAA). Such support documentation is appended herewith as Exhibit B.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Date: 5-14-06



John G. Babish, Ph.D.
Executive Vice President
Metagenics Research Center - Suite 100
9770 44th Ave. NW
Gig Harbor, WA 98332

FILED: 5/14/2006 10:00:00

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Exhibit A**BIOGRAPHICAL SKETCH AND BIBLIOGRAPHY**

John G. Babish

Chairperson, BIONexus, Ltd.
Executive Vice President, Metaproteomics Inc.**Education**

Institution and Location of Study	Degree	Date Conferred	Field
The Pennsylvania State University, State College, PA	B.S	1968	Biochemistry
Cornell University, Ithaca, NY	M.S	1974	Chemistry
Cornell University, Ithaca, NY	Ph.D.	1976	Biochemistry

Research and Professional Experience

Aug. 2002 – present Executive Vice President of Research & Development, Metaproteomics, Research Laboratories, Ithaca, NY. Metaproteomics develops clinically proven, patented dietary supplements and pharmaceuticals from natural sources. Duties include the design and evaluation of experiments elucidating mechanism of action and biological activity within complex mixtures.

1998 – present (5% Effort) National Coordinator for the USDA Minor Species Drug Program (NRSP-7). The NRSP-7 program is funded by the USDA to provide funds and expertise necessary for the approval of pharmaceuticals used in the treatment of diseases associated with minor crop species. Duties include the coordination of industrial, academic and regulatory resources necessary for protocol development through final drug approval.

1997 – present Co-founder and Chairperson of BIONexus, Ltd. Ithaca, NY. BIONexus develops and markets nutritional supplements to address health problems associated with AIDS. NutriVir™, the BIONexus supplement for wasting in HIV/AIDS, generated approximately \$600,000 in gross revenues in its first year of sales. NutriVir™ is reimbursed by Medicaid in 14 states.

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- 1991 – 1996 Founder, Chairperson, President and CEO of Paracelsian, Inc., Ithaca, NY. The Company was launched from the technology transfer program of Cornell University in 1991, and with the public offering in 1992 (Nasdaq:PRLN), became the first public corporation of a Cornell University technology transfer effort. Babish was associated with the attainment of over \$12 million dollars in corporate financing.
- 1984 – 1996 Tenured, Associate and Professor of Pharmacology and Toxicology, Department of Pharmacology, College of Veterinary Medicine, Cornell University. Offered the first course in molecular risk assessment in the USA in 1979; member of the graduate Fields of Pharmacology, Toxicology, Veterinary Medicine, Food Science and Epidemiology; successfully petitioned the State of New York for the approval of the separate Fields of Toxicology and Pharmacology at Cornell University.
- 1978 – 1984 Assistant Professor, Department of Preventive Medicine, NYS College of Veterinary Medicine, Cornell University, Ithaca, NY.
- 1976 – 1978 Postdoctoral Scientist, Food and Drug Research Labs, Waverly, NY.

Invited Presentations (Recent of 38 presentations)

Micronutrient deficiencies in AIDS wasting at Progressive Management of AIDS Wasting: 2000. Hunter College, NYC. March 24, 2000.

Phytochemicals and NF- κ B activation at IBC's Conference on The Health Benefits of Natural Phytochemicals. Montreal Bonaventure Hilton, July 22 – 23, 1997.

Chemically-induced cell cycle stasis in immunotoxicology. 12th Annual NIOSH Conference on Mechanisms of Immunotoxicology – Role of Apoptosis in Immunotoxicology. University of West Virginia, Morgantown, WV. September 10 – 12, 1997.

Publications (Selected of 108 peer-reviewed publications)

Payne M.A., Babish J.G., Bulgin M., Lane M., Wetzlich S., Craigmill A.L. (2002) Serum pharmacokinetics and tissue and milk residues of oxytetracycline in goats following a single intramuscular injection of a long-acting preparation and milk residues following a single subcutaneous injection. *J Vet Pharmacol Ther.* 25(1):25-32.

Calabrese C., Berman S.H., Babish J.G., et al. (2000) A phase I trial of andrographolide in HIV positive patients and normal volunteers. *Phytother Res.* 14(5):333-338.

Ma, X., Stoffregen, D.A., Wheelock, G.D., Rininger, J.A. and Babish, J.G. (1997) Discordant hepatic expression of the cell division control enzyme p34cdc2 kinase, proliferating cell nuclear antigen, p53 tumor suppressor protein, and p21Waf1 cyclin-

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dependent kinase inhibitory protein after WY14,643 ([4-chloro-6-(2,3-xylidino)-2-pyrimidinylthio]acetic acid) dosing to rats. *Mol. Pharmacol.*, 51, 69-78.

Rininger, J.A., Goldsworthy, T.L. and Babish, J.G. (1997) Time course comparison of cell-cycle protein expression following partial hepatectomy and WY14,643-induced hepatic cell proliferation in F344 rats. *Carcinogenesis*, 18, 935-941.

Rininger, J.A., Stoffregen, D.A. and Babish, J.G. (1997) Murine hepatic p53, RB, and CDK inhibitory protein expression following acute 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) exposure. *Chemosphere*, 34, 1557-1568.

Rininger, J.A., Wheelock, G.D., Ma, X. and Babish, J.G. (1996) Discordant expression of the cyclin-dependent kinases and cyclins in rat liver following acute administration of the hepatocarcinogen [4-chloro-6-(2,3-xylidino)-2-pyrimidinylthio] acetic acid (WY14,643). *Biochem. Pharmacol.*, 52, 1749-1755.

Vancutsem, P.M. and Babish, J.G. (1996) In vitro and in vivo study of the effects of enrofloxacin on hepatic cytochrome P-450. Potential for drug interactions. *Vet. Hum. Toxicol.*, 38, 254-259.

Patents (Selected of 15 US and three foreign patents)

US Patent No. 5,833,994	11/10/1998 Use of the Ah receptor and Ah receptor ligands to treat or prevent cytopathicity of viral infection.
US Patent No. 5,612,188	3/18/1997 Automated, multicompartmental cell culture system.
US Patent No. 5,529,899	6/25/1996 Immunoassay for Ah receptor transformed by dioxin-like compounds.
US Patent No. 5,496,703	3/5/1996 Indirect immunoassay for dioxin-like compounds

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Exhibit B

LABORATORY NOTEBOOK SHEETS DOCUMENTING RESEARCH ON RIAA, IAA AND OTHER HOPS
DERIVATIVES

Date	Notebook Number	Pages
6/4/02 - 8/24/02	2002-03	13 - 23
8/28/02	2002-04	3 - 4
12/5/02	2002-06	1 - 2
3/5/03	2002-08	43 - 44
4/23/03	2003-01	23
6/5/03	2003-01	45
9/3/03	2003-4	22
12/15/03	2003-5	42

PROJECT POE₂ Assay in RAW cellsNotebook No. 2002-03

Continued From Page

13

Experiment 2002-03-13

The general purpose + procedure can be found on pages 4, 8, and 11 of this notebook

1_C00X-2RAW_004.02

Compounds for Metagenics
POC2 assay, using 1:10 dilution from the plates
6.04.02 (USING RAW 264.7 cells with LPS stimulation and 15 min with ARA)

Compound	Conc	100µl	10µl	1µl	0.1µl	0.01µl
1. BetaTech - Alpha hop (10µg/L)	Alpha hop =	50,000	10,000	5,000	1,000	0
2. BetaTech - Beta acid solution (10µg/L)	Beta acid solution =	50,000	10,000	5,000	1,000	0
3. BetaTech Aromashop OE (10µg/L)	Aromashop OE =	50,000	10,000	5,000	1,000	0
4. BetaTech Radshop (RAA) (10µg/L)	Radshop (RAA) =	50,000	10,000	5,000	1,000	0
5. BetaTech Radshop (RIAA) (10µg/L)	Radshop (RIAA) =	50,000	10,000	5,000	1,000	0
6. BetaTech Tetrahop Gold (10µg/L)	Tetrahop Gold =	50,000	10,000	5,000	1,000	0
7. BetaTech Hexahop gold (HRAA) (10µg/L)	Hexahop gold =	50,000	10,000	5,000	1,000	0
8. BetaTech Hop oil (10µg/L)	Hop oil =	50,000	10,000	5,000	1,000	0
9. Dibutylpropyl fluorophosphate	DIPP =	50,000	5,000	0,500	0,050	0
10. Indinavir HCl Tablets	Indinavir HCl =	50,000	25,000	12,500	6,250	80

2_C00X-2RAW_004.02

New Hops Compounds for Metagenics
POC2 assay, using 1:10 dilution from the plates
6.04.02 (USING RAW 264.7 cells with LPS stimulation and 15 min with ARA)

Compound	Conc	100µl	10µl	1µl	0.1µl	0.01µl
1. BetaTech Radshop (RAA) (10µg/L)	RAA =	50,000	10,000	5,000	1,000	0
2. RAA/uronic acid (1:1)	Total = RAA = Uronic acid =	50,000 25,000 25,000	10,000 5,000 5,000	5,000 2,500 2,500	1,000 0,500 0,500	0
3. Lemon Bioflavonoid (RMD77810)	Bioflavonoid =	50,000	10,000	5,000	1,000	0
4. Ginger (RMD7782)	Ginger =	50,000	10,000	5,000	1,000	0
5. RAA/Cucurbitin (RMD7782) (1:1)	Total = RAA = Cucurbitin =	50,000 25,000 25,000	10,000 5,000 5,000	5,000 2,500 2,500	1,000 0,500 0,500	0
6. Capsaicin Pepper (RMD7782)	Capsaicin =	50,000	10,000	5,000	1,000	0
7. RAA/Quercetin (RMD7781) (1:1)	Total = RAA = Quercetin =	50,000 25,000 25,000	10,000 5,000 5,000	5,000 2,500 2,500	1,000 0,500 0,500	0
8. RAA/Silphib gelatin (1:1)	Total = RAA = Silphib gelatin =	50,000 25,000 25,000	10,000 5,000 5,000	5,000 2,500 2,500	1,000 0,500 0,500	0
9. RAA/Boswellin (RMD7781) (1:1)	Total = RAA = Boswellin =	50,000 25,000 25,000	10,000 5,000 5,000	5,000 2,500 2,500	1,000 0,500 0,500	0
10. Henna/oxide	Henna/oxide =	50,000	10,000	5,000	1,000	0

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Date

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Date

PROJECT

BEJ In RAW cells

Notebook No 2002-03

Continued From Page

16

Experiment 2002-03 -16

The purpose and procedure for this experiment can be found on pages 4, 8, and 16 of this notebook.

3_COX-2RAW_8.10.02

Compounds for Metagenesis
PGE2 assay, using 1:10 dilution from the plates
8.10.02 USING RAW 264.7 cells with LPS stimulation and 15 min with ARAI. Data media 1:50 and 1:100 for PGE2 assay

1. BetaTech - Alpha hop (10µg/dL)	Alpha hop =	50,000	10,000	5,000	1,000	0
2. BetaTech - Beta acid solution (10µg/dL)	Beta acid solution =	50,000	10,000	5,000	1,000	0
3. BetaTech Aromahop OE (10µg/dL)	Aromahop OE =	50,000	10,000	5,000	1,000	0
4. BetaTech Isachop (IAA) (10µg/dL)	Isachop (IAA) =	50,000	10,000	5,000	1,000	0
5. BetaTech Radhop (RAA) (10µg/dL)	Radhop (RAA) =	50,000	10,000	5,000	1,000	0
6. BetaTech Tetrahop Gold (10µg/dL)	Tetrahop Gold =	50,000	10,000	5,000	1,000	0
7. BetaTech Hexahop gold (HMAA) (10µg/dL)	Hexahop gold =	50,000	10,000	5,000	1,000	0
8. Ibuprofen	Ibuprofen =	50,000	10,000	5,000	1,000	0
9. Celebrex	Celebrex =	5,000	0,500	0,050	0,005	0
10. Infraredoid IC	Infraredoid IC =	50,000	10,000	5,000	1,000	0
					total =	80

BIO-TEK MICROPLATE READER 06/14/02 AT 01:02 PM 00011864

ASSAY

PLATE

OPERATOR

NOTES

PROGRAM MODE #9

SINGLE WAVELENGTH: 495

TABLE OF ABSORBANCE VALUES

	1	2	3	4	5	6	7	8	9	10	11	12
A	0.697	0.439	0.391	0.341	0.376	0.317	0.329	0.346	0.342	0.697	0.667	0.678
B	1.028	0.333	0.366	0.323	0.371	0.333	0.321	0.326	0.344	0.683	0.609	0.704
C	0.193	0.666	0.319	0.382	0.479	0.313	0.334	0.348	0.384	0.397	0.662	0.398
D	0.163	0.637	0.333	0.314	0.316	0.331	0.327	0.363	0.321	0.309	0.659	0.388
E	0.368	0.378	0.399	0.377	0.362	0.381	0.376	0.616	0.638	0.615	0.649	0.373
F	0.667	0.386	0.314	0.313	0.333	0.334	0.337	0.336	0.326	0.352	0.648	0.354
G	0.136	0.673	0.323	0.332	0.344	0.346	0.388	0.349	0.333	0.396	0.684	0.682
H	0.103	0.653	0.319	0.373	0.319	0.347	0.317	0.348	0.346	0.331	0.638	0.534

BIO-TEK MICROPLATE READER 06/14/02 AT 01:03 PM 00011865

ASSAY

PLATE

OPERATOR

NOTES

PROGRAM MODE #9

SINGLE WAVELENGTH: 495

TABLE OF ABSORBANCE VALUES

	1	2	3	4	5	6	7	8	9	10	11	12
A	0.988	0.641	0.682	0.681	0.639	0.399	0.628	0.621	0.627	0.708	0.694	0.728
B	1.081	0.628	0.333	0.378	0.628	0.682	0.394	0.685	0.624	0.698	0.782	0.732
C	0.168	0.639	0.366	0.333	0.331	0.333	0.386	0.378	0.398	0.638	0.664	0.657
D	0.163	0.637	0.384	0.331	0.372	0.377	0.383	0.606	0.398	0.612	0.678	0.633
E	0.678	0.487	0.633	0.612	0.617	0.648	0.623	0.626	0.398	0.649	0.663	0.661
F	0.393	0.396	0.389	0.383	0.364	0.398	0.377	0.689	0.682	0.628	0.658	0.622
G	0.134	0.673	0.623	0.398	0.388	0.373	0.396	0.384	0.638	0.631	0.783	0.662
H	0.132	0.688	0.646	0.648	0.628	0.631	0.638	0.633	0.782	0.616	0.783	0.628

The same plate was run @ 2 different dilutions. 1:50 and 1:100

Continued on Page

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Signed

Date

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Date

PROJECT POE Assay + NO AssayNotebook No. 2002-03

Continued From Page

17

Experiment 2002-03-17

- The general purpose and procedure for the POE Assay can be found on 2002-03-04 experiment.
- Purpose for the NO experiment to determine the levels of Nitrite Oxide being produced in the cells.

- Procedure for NO

Griess Reagent was made.

1% - Sulfanilamide Sigma 5-9251 lot 101K2010
 0.1% - N-(1-naphthyl) ethylenediamine (Sigma N-5889)
 2.5% - H₃PO₄ (Riedel-de Haen -0410) lot 71150112
 Lot 52540

This reagent is mixed up in Ultra Pure Water

25 ml = 250 mg Sulfanilamide
 25 mg N-(1-naphthyl) ethylenediamine
 625 mg H₃PO₄

The same plates of cells were used for each experiment.

The cells were taken from 1 X T₇₅ plate of RAW cells that were 80% confluent and plated into 2 96 well plates at 4×10^5 cells per well.

Continued on Page 18

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[Signature] 6-17-02
 Signed Date

[Signature] 7-19-02
 Signed Date

PROJECT PGC₂ + NO in RAN cellsNotebook No. 2002-03Continued From Page 17

18

2002-03-15/17 cont.

1. CON-RAW_6.17.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates

6.17.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 8 μM ARA1

Compound	50.000	10.000	5.000	1.000	8
1. BetaTech - Alpha hop (10μg/L)	Alpha hop = 50.000	10.000	5.000	1.000	8
2. BetaTech - Beta acid solution (10μg/L)	Beta acid solution = 50.000	10.000	5.000	1.000	8
3. BetaTech Aromashop OE (10μg/L)	Aromashop OE = 50.000	10.000	5.000	1.000	8
4. BetaTech Isoshop (IAA) (10μg/L)	Isoshop (IAA) = 50.000	10.000	5.000	1.000	8
5. BetaTech Radshop (RIAA) (10μg/L)	Radshop (RIAA) = 50.000	10.000	5.000	1.000	8
6. BetaTech Tetrahop Gold (10μg/L)	Tetrahop Gold = 50.000	10.000	5.000	1.000	8
7. BetaTech Hexahop gold (HIAA) (10μg/L)	Hexahop gold = 50.000	10.000	5.000	1.000	8
8. Ibuprofen	Ibuprofen = 50.000	10.000	5.000	1.000	8
9. Celebrex	Celebrex = 5.000	0.500	0.050	0.005	8
10. Inflamoid IC Tablets	Inflamoid IC = 50.000	10.000	5.000	1.000	8
				total =	80

The compounds used on these two plates are identical. The amount of Arachidonic Acid used is not.

4 plates will be used for PGE₂ and

Three for NO

2. CON-RAW_6.17.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates

6.17.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 8 μM ARA1

Compound	50.000	10.000	5.000	1.000	8
1. BetaTech - Alpha hop (10μg/L)	Alpha hop = 50.000	10.000	5.000	1.000	8
2. BetaTech - Beta acid solution (10μg/L)	Beta acid solution = 50.000	10.000	5.000	1.000	8
3. BetaTech Aromashop OE (10μg/L)	Aromashop OE = 50.000	10.000	5.000	1.000	8
4. BetaTech Isoshop (IAA) (10μg/L)	Isoshop (IAA) = 50.000	10.000	5.000	1.000	8
5. BetaTech Radshop (RIAA) (10μg/L)	Radshop (RIAA) = 50.000	10.000	5.000	1.000	8
6. BetaTech Tetrahop Gold (10μg/L)	Tetrahop Gold = 50.000	10.000	5.000	1.000	8
7. BetaTech Hexahop gold (HIAA) (10μg/L)	Hexahop gold = 50.000	10.000	5.000	1.000	8
8. Ibuprofen	Ibuprofen = 50.000	10.000	5.000	1.000	8
9. Celebrex	Celebrex = 5.000	0.500	0.050	0.005	8
10. Inflamoid IC Tablets	Inflamoid IC = 50.000	10.000	5.000	1.000	8
				total =	80

- 1 - Alpha hop - AN1124
 2 - Beta Acid - AN1125
 3 - Aromashop - AN1126
 4 - Isoshop - AN1127
 5 - Radshop - AN1128
 6 - Tetrahop - AN1128
 7 - Hexahop - AN1129
 8 - Ibuprofen - Sigma I-4683 lot 26 H1368
 9 - Celebrex - AN1055
 10 - Inflamoid IC Tablets - AN1101

Continued on Page 19

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6.19.02
6.15.02
 Date

[Signature]
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7-19-02
 Date

PROJECT BE₂ + NO Assay in RAW cells.

Notebook No. 2002-03-4974
Continued From Page 18

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Experiment 2002-03-17 cont.

~~The MO plates were in the~~

The plates were treated and supernate collected for the PGE₂ plots. Then the plates were put back into the incubator overnight.

GOX-2 1.2

فہمنا ہا عتہ

NSA-Tek Data Summary

Wesley, Duick Reed

Date: 06/20/02

Not,

Time: 02:30:53 AM

Opinion Editor

Temp 1
14864415

Place for

PC 100075

[illegible]

This is the raw data for the Co^{+2} plate diluted 1:2

This is the R.A. data for the co-2 plate diluted 1:10.

b6-Denkmal 2025-08-22 14:00:00

Answer: Quilok Road

Date: 06/30/07

Lot.

Time: 02:32:20 PM

Operations

११५००

Place in
148344124

CONTACT:

[illegible]

Continued on Page 70

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Signed

6.70.02

Date _____

Signed _____

Signed

7-19-02

Date _____

PROJECT PGE₂ + NO₃ in RAW cellsNotebook No. 2002-03
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Experiment 2002-03-17 cont.

COX-1 1.2

Bio-Tek Instruments

Assay: Quick Read

Date: 06/30/03

Lot:

Wavelength: 405

Time: 07:35:43PM

Operator:

Assay: 405

Temp:

Plate ID:

Comments:

This is the data from
the RAW plate COX-1
1:2 dilution.

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.140	0.130	0.130	0.147	0.177	0.157	0.160	0.130	0.130	0.130	0.130
Well	001	002	003	004	005	006	007	008	009	010	011	012
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	013	014	015	016	017	018	019	020	021	022	023	024
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	025	026	027	028	029	030	031	032	033	034	035	036
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	037	038	039	040	041	042	043	044	045	046	047	048
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	049	050	051	052	053	054	055	056	057	058	059	060
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	061	062	063	064	065	066	067	068	069	070	071	072
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	073	074	075	076	077	078	079	080	081	082	083	084
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140	0.140
Well	085	086	087	088	089	090	091	092	093	094	095	096
Rep												

This is data from the
Undiluted COX-1 plate.

Assay: Quick Read

Date: 06/30/03

Lot:

Wavelength: 405

Time: 07:34:04PM

Operator:

Assay: 405

Temp:

Plate ID:

Comments:

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	001	002	003	004	005	006	007	008	009	010	011	012
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	013	014	015	016	017	018	019	020	021	022	023	024
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	025	026	027	028	029	030	031	032	033	034	035	036
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	037	038	039	040	041	042	043	044	045	046	047	048
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	049	050	051	052	053	054	055	056	057	058	059	060
Rep												

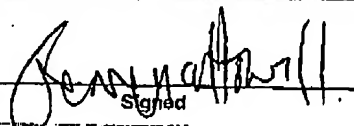
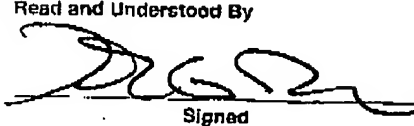
	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	061	062	063	064	065	066	067	068	069	070	071	072
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	073	074	075	076	077	078	079	080	081	082	083	084
Rep												

	1	2	3	4	5	6	7	8	9	10	11	12
Cell												
Optical Density	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130	0.130
Well	085	086	087	088	089	090	091	092	093	094	095	096
Rep												

Continued on Page 21

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 Date 6-20-07

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 Date 7-19-02

PROJECT PGE₂ + NO in RAW cells

Notebook No. 780203
Continued From Page 20

Continued From Page 10

21

Experiment 2002-03-17 cont

A standard curve was done on the NO.

50ul of Media Δ of the treated cells
was added to 50ul of the Greiss reagent
This was allowed to sit for 10 minutes
then read @ 605 nm.

No St. Curve

Abstract

ANALYST: CLYDE R. BARNES

Date: 06/20/02

1051

wordlength: 630

Time: 02:28:12.99
 Date: 1

Operator:
Plate ID:

1116-224

0000000000

This is the 10
Standard curve

[illegible]

Continued on Page 22

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Date

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Date _____

PROJECT REG - No Assay in RAW cellsNotebook No. 2002-03Continued From Page 21

22

Experiments 2002-03-17

Bio-Tek Instruments

Assay: Quick Read

Date: 08/20/02

Lot:

Assay length: 400

Time: 02:33:40 PM

Operator:

Temp: 37.0

Plate ID:

CONCENTRATIONS

COX-2 RAW data
NO

Assay: Quick Read

Date: 08/20/02

Lot:

Assay length: 400

Time: 02:38:07 PM

Operator:

Temp: 37.0

Plate ID:

CONCENTRATIONS

COX-1 RAW data
NO

Continued on Page

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Date

Signed

Date

PROJECT PGC₂ in RAW cells + NO AssayNotebook No. 2002-03

Continued From Page

23

Experiment 2002-03-23

The purpose and procedure can be found on page 11 of this notebook.

1_COX-2RAW_8.24.02

Compounds for Metagenics

PGC₂ assay, using 1:1 and 1:10 dilution from the plates

6.24.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 8 µM ARA

Compound	Concentration	50.000	5.000	0.500	0.050	8
1. Alcohol extract of spent hops	EXHSH =	50.000	5.000	0.500	0.050	8
2. Urolic acid 90% - Salinas	Urolic acid =	50.000	5.000	0.500	0.050	8
3. Cinnamic acid (Rosemary)	Salutic acid =	50.000	5.000	0.500	0.050	8
4. Cinnamic acid 80% - Salinas	Cinnamic acid 80% =	50.000	5.000	0.500	0.050	8
5. BetaTech Radtop (RAA) (10 µg/L)	Radtop (RAA) =	50.000	5.000	0.500	0.050	8
6. Curcumin granular (Q856)	Curcumin =	50.000	5.000	0.500	0.050	8
7. RAA:Urolic acid (90%) - (1:1)	Total =	50.000	5.000	0.500	0.050	8
	RAA =	25.000	2.500	0.250	0.025	
	Urolic acid 90% =	25.000	2.500	0.250	0.025	
8. Ibuprofen - Sigma	Ibuprofen =	50.000	5.000	0.500	0.050	8
9. Celebrex	Celebrex =	5.000	0.500	0.050	0.005	8
10. Aspirin - Sigma	Aspirin =	50.000	10.000	5.000	1.000	8
					total =	80

2_COX-2RAW_8.24.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates

6.24.02 USING RAW 264.7 cells no LPS stimulation and 60 min with 100 µM ARA

Compound	Concentration	50.000	5.000	0.500	0.050	8
1. Alcohol extract of spent hops		50.000	5.000	0.500	0.050	8
2. Urolic acid 90% - Salinas		50.000	5.000	0.500	0.050	8
3. Cinnamic acid (Rosemary)		50.000	5.000	0.500	0.050	8
4. Cinnamic acid 80% - Salinas		50.000	5.000	0.500	0.050	8
5. BetaTech Radtop (RAA) (10 µg/L)	Radtop (RAA) =	50.000	5.000	0.500	0.050	8
6. Curcumin granular (Q856)		50.000	5.000	0.500	0.050	8
7. RAA:Urolic acid (90%) - (1:1)	Total =	50.000	5.000	0.500	0.050	8
	RAA =	25.000	2.500	0.250	0.025	
	Urolic acid 90% =	25.000	2.500	0.250	0.025	
8. Ibuprofen - Sigma	Ibuprofen =	50.000	5.000	0.500	0.050	8
9. Celebrex	Celebrex =	5.000	0.500	0.050	0.005	8
10. Aspirin - Sigma	Aspirin =	50.000	10.000	5.000	1.000	8
					total =	80

Continued on Page 24

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Signed

6.24.02

Date



Signed

7-19-02

Date

PROJECT

PGE₂ Expression in AGS + A-549 cells

Notebook No.

Continued From Page

3

2002-0304-01 cont.

C2_COX2_A549_8.22.02

A549 Cells - wash, cells treated with test material, stimulated with IL-1/LPS/IL-6 for 24 hrs and assayed for PGE₂.
For PGE₂ assay run media undiluted and diluted 1:20.

1. DataTech (IAA)	IAA =	25	5.0	0.5	0.05	0
2. DataTech (RIA)	DataTech (RIA) =	25	5.0	0.5	0.05	0
3. IAA:RIA (1:1)	Total =	25	5.0	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	RIA =	12.5	2.5	0.25	0.025	0
4. IAA:RIA (2:1)	Total =	25	5	0.5	0.05	0
	IAA =	16.7	3.3	0.33	0.033	0
	RIA =	8.3	1.7	0.167	0.017	0
5. IAA:RIA (5:1)	Total =	25	5	0.5	0.05	0
	IAA =	20.0	4.0	0.4	0.04	0
	RIA =	4.2	0.8	0.08	0.008	0
6. IAA:RIA (10:1)	Total =	25	5	0.5	0.05	0
	IAA =	22.7	4.55	0.455	0.0455	0
	RIA =	2.3	0.45	0.045	0.0045	0
7. IAA:RIA (50:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.0	4.80	0.480	0.0480	0
	RIA =	0.5	0.10	0.010	0.001	0
8. IAA:RIA (100:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.75	4.95	0.495	0.0495	0
	RIA =	0.25	0.05	0.0050	0.0005	0
9. IAA:TrypanBlue (1:1)	Total =	25	5	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	TrypanBlue =	12.5	2.5	0.25	0.025	0
Total =						72

Note 1000, 500, 100 and 50 concentrations of the standard curve are listed in Column 2

This plate was treated the same as the plate on page 2 of this notebook

C3_COX2_A549_8.22.02B6

A549 Cells - cells are stimulated with IL-1/LPS/IL-6 for 24 hrs, washed, test materials added for 60 minutes then A23187 (50 µM) added as assay for PGE₂ 30 minutes later.
For PGE₂ assay run media undiluted and 1:20.

1. Diisopropyl fluorophosphate	COX-2	25	5.0	0.5	0.05	0
2. Vioxx	COX-2	25	5.0	0.5	0.05	0
3. Celecoxib	COX-2	25	5.0	0.5	0.05	0
4. Nimesulide	COX-2	25	5.0	0.5	0.05	0
5. Ibuprofen	COX-2/COX-1	25	5.0	0.5	0.05	0
6. Indomethacin	COX-1	25	5.0	0.5	0.05	0
7. Aspirin	COX-1	25	5.0	0.5	0.05	0
8. Salicylic acid	-	25	5.0	0.5	0.05	0
9. Naproxen	-	25	5.0	0.5	0.05	0
10. Acetaminophen	-	25	5.0	0.5	0.05	0
Total =						80

- 1- Aldrich D12/600 lot 0245625
- 2- AN1066
- 3- AN1055
- 4- N-1016 lot 117H1019
- 5- Sigma I-4863 lot 26H1268
- 6- Sigma I-7578 lot 60K0745
- 7- A-5376 lot 119H0175
- 8- AN1071
- 9- N-8280 lot 11K1767
- 10- A-2085 lot 20K068
- A-23187 - Sigma
- C-7522 lot 81K4002

This plate was stimulated with IL-1/LPS and H₂O₂ for 24 hrs. The plate will then be washed and test materials added for 1 hr then stimulated with A23187 for 30 minutes.

Continued on Page 4

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Date

8-28-02

[Signature]
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Date

9-1-02

PROJECT POE₂ Expression in A-549 CellsNotebook No. 2002-04Continued From Page 3

AGS Cells Format A - wash, add test material and assay for POE₂ the next day
For POE₂ assay run media undiluted
8.34.02

Experiment 2002-04-01

1. Oleic acid (50% Oleic acid)	Oleic acid =	25	5.0	0.5	0.05	0
2. BetaTech (RIAA)	BetaTech (RIAA) =	25	3.0	0.3	0.05	0
3. TrypanBlue - Water Channels	TrypanBlue =	10	1	0.1	0.01	0
4. RIAA:Oleic acid - (1:1)	Total =	25	5	0.5	0.05	0
	RIAA =	22.7	4.5	0.453	0.045	0
	Oleic acid =	2.3	0.5	0.045	0.005	0
5. RIAA:Oleic acid - (5:1)	Total =	25	5	0.5	0.05	0
	RIAA =	20.0	4.2	0.417	0.042	0
	Oleic acid =	4.2	0.8	0.083	0.008	0
6. RIAA:Oleic acid - (1:5)	Total =	25	5	0.5	0.05	0
	RIAA =					
	Oleic acid =					
7. RIAA:Oleic acid - (1:10)	Total =	25	5	0.5	0.05	0
	RIAA =					
	Oleic acid =					
8. #1115 Metoprolol	#1115 =	25	5	0.5	0.05	0
9. RIAA:TrypanBlue - (1:1)	Total =	25	5	0.5	0.05	0
	RIAA =					
	TrypanBlue =					
		Total =	72			

The first two plates were run in the ~~exact~~ same manner as the A-549 plates outlined on pages 2 and three of this notebook

Run 1000, 500, 15.0 and 7.5 concentrations of the standard curve as series in Column 2

AGS Cells Format A - wash, add test material and assay for POE₂ the next day
For POE₂ assay run media undiluted
8.34.02

1. BetaTech (RIAA)	RIAA =	25	5.0	0.5	0.05	0
2. BetaTech (RIAA)	BetaTech (RIAA) =	25	5.0	0.5	0.05	0
3. IAA:RIAA - (1:1)	Total =	25	5.0	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	RIAA =	12.5	2.5	0.25	0.025	0
4. IAA:RIAA - (2:1)	Total =	25	5	0.5	0.05	0
	IAA =	16.7	3.3	0.333	0.033	0
	RIAA =	8.3	1.7	0.167	0.017	0
5. IAA:RIAA - (5:1)	Total =	25	5	0.5	0.05	0
	IAA =	20.0	4.0	0.4	0.04	0
	RIAA =	4.2	0.8	0.08	0.008	0
6. IAA:RIAA - (10:1)	Total =	25	5	0.5	0.05	0
	IAA =	22.7	4.5	0.453	0.045	0
	RIAA =	2.3	0.5	0.045	0.005	0
7. IAA:RIAA - (20:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.5	4.9	0.49	0.049	0
	RIAA =	0.5	0.1	0.01	0.001	0
8. IAA:RIAA - (100:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.75	4.95	0.495	0.0495	0
	RIAA =	0.25	0.05	0.005	0.0005	0
9. IAA:TrypanBlue - (1:1)	Total =	25	5	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	TrypanBlue =	12.5	2.5	0.25	0.025	0
		Total =	72			

Run 1000, 500, 15.0 and 7.5 concentrations of the standard curve as series in Column 2

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 µM) added and assay for POE₂ 30 minutes later.
For POE₂ assay run media undiluted and 120
8.34.02

1. Diisopropyl fluorophosphate	COX-2	25	5.0	0.5	0.05	0
2. Water	COX-2	25	5.0	0.5	0.05	0
3. Celecoxib	COX-2	25	5.0	0.5	0.05	0
4. Nimodipine	COX-2	25	5.0	0.5	0.05	0
5. Ibuprofen	COX-2/COX-1	25	5.0	0.5	0.05	0
6. Indomethacin	COX-1	25	5.0	0.5	0.05	0
7. Aspirin	COX-1	15	5.0	0.5	0.05	0
8. Salicylic acid	-	25	5.0	0.5	0.05	0
9. Naproxen	-	25	5.0	0.5	0.05	0
10. Acetaminophen	-	25	5.0	0.5	0.05	0
		Total =	80			

Plate 3 was done with the test materials added and then 60 minutes later they were stimulated w/ A23187 for 30 minutes before samples were taken for POE₂ plates

Continued on Page 5

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Date

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Date

PROJECT Cell Lyse + Keko PUG 37Notebook No 2002-06
Continued From Page 1

2

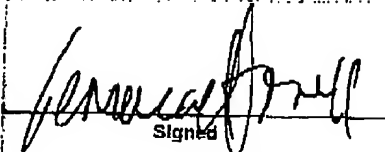
Experiment + 2002-06-01

COX-2 - 56KD

COX-1 66KD

iNOS 130KD

Continued on Page


Signed12-5-02
Date

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Signed12-23-02
Date

PROJECT

PGE Assay in AGS cells

Notebook No. 2002-06

Continued From Page

43

Experiment 2002-06-43

Purpose + Procedure can be found on pages 2002-06-11

C1_COXAGS_03.03.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 μ M) added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
3.03.03

Compound	Comments	d1 (μ g/mL)	d2 (μ g/mL)	d3 (μ g/mL)	d4 (μ g/mL)	No Wells
1. Uridine diphosphate		50	5.0	0.5	0.05	8
2. METABOL		50	5.0	0.5	0.05	8
3. Rosemary extract (07720)		50	5.0	0.5	0.05	8
4. Quercetin acid (50% Gabassa)		50	5.0	0.5	0.05	8
5. RIAA (Hase)		50	5.0	0.5	0.05	8
6. Rutin (08294)		50	5.0	0.5	0.05	8
7. Curcumin (07387)		50	5.0	0.5	0.05	8
8. Ginger root (06836)		50	5.0	0.5	0.05	8
9. AHEB		50	5.0	0.5	0.05	8
10. Aspirin (Stam)		50	5.0	0.5	0.05	8
Total =						80

C2_COXAGS_03.03.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 μ M) added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
3.03.03

Compound	Comments	d1 (μ g/mL)	d2 (μ g/mL)	d3 (μ g/mL)	d4 (μ g/mL)	No Wells
1. Isobut AN1127		50	5.0	0.5	0.05	8
2. B1115		50	5.0	0.5	0.05	8
3. Tolshop AN1120		50	5.0	0.5	0.05	8
4. Hmshop AN1130		50	5.0	0.5	0.05	8
5. Alphashop AN1124		50	5.0	0.5	0.05	8
6. BetaShop AN1126		50	5.0	0.5	0.05	8
7. Iso-Rich AN1060		50	5.0	0.5	0.05	8
8. Tannin Extract #411 AN1173		50	5.0	0.5	0.05	8
9. A70 LIPOTECH		50	5.0	0.5	0.05	8
10. Aromashop AN1120		50	5.0	0.5	0.05	8
Total =						80

Continued on Page 44

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Date

PROJECT PCE, Army in ASA calls

Notebook No. 2002-05

Continued From Page 43

44

Experiment - 2002-6-03 con +

After Def.

NSA-TR-2010-01

Assay: Quick Read

Date: 05/07/05

Lot. 16 S 1:20 Plate 1

0177472:465

2600 112:47:57800

929596

Plata 7D

Summary:

[illegible]

Die-Tek Instruments

Essay: Quiet Read

DATA: 01/07/02

tot: 196 1140

valued at 409

Temp: _____

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11/11/2019

CONCLUSIONS

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3-2-03
Date

Date


Sinner

3-9-03

Date _____

PROJECT AGE₂ Assay in AGS cellsNotebook No. 2003-01

Continued From Page

23

Experiment 2003-01-23

Purpose + Procedure: The purpose is to look at the expression of AGE₂ in AGS cells when exposed to multiple natural compounds.

The procedure can be found on 2003-01-23

4.23.03 - The cells were seeded at 7x10⁵ cells per well in 2-96 well plates.

The following treatments will be used on the AGS cells: AGS cells - ATCC # HTB-79, lot 1641648

AGS Cells - cells grown to confluence, wash, feed medium added for 30 minutes then AGE₂ (50 µM) added and assay for AGE₂ 24 minutes later.
For AGE₂ assay run media diluted 1:20 only
5.23.03

Compound	Comments	d1 [µg/mL]	d2 [µg/mL]	d3 [µg/mL]	d4 [µg/mL]	No Wells
1. AGE ₂ (100%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
2. AGE ₂ (50%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
3. AGE ₂ (10%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
4. AGE ₂ (1%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
5. AGE ₂ (0.1%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
6. AGE ₂ (0.01%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
7. AGE ₂ (0.001%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
8. AGE ₂ (0.0001%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
9. AGE ₂ (0.00001%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
10. AGE ₂ (0.000001%)	Compound = 50 RNA = 25 Exposure = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		

C1_COXAGS_04.21.03

AGS Cells - cells grown to confluence, wash, feed medium added for 30 minutes then AGE₂ (50 µM) added and assay for AGE₂ 24 minutes later.
For AGE₂ assay run media diluted 1:20 only
5.23.03

Compound	Comments	d1 [µg/mL]	d2 [µg/mL]	d3 [µg/mL]	d4 [µg/mL]	No Wells
1. Isohex AN1127		50	5.0	0.5	0.05	8
2. 61116		50	5.0	0.5	0.05	8
3. Tetrahex AN1120		50	5.0	0.5	0.05	8
4. Hexahept AN1130		50	5.0	0.5	0.05	8
5. Alphahept AN1124		50	5.0	0.5	0.05	8
6. Betahept AN1128		50	5.0	0.5	0.05	8
7. Iso-Rich AN1090		50	5.0	0.5	0.05	8
8. Tarron Extract 84411 AN1173		50	5.0	0.5	0.05	8
9. AYU LIPOTECH		50	5.0	0.5	0.05	8
10. Aromahex AN1125		50	5.0	0.5	0.05	8
Total =						80

Continued on Page 22

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Jenniforwell
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4.23.03

Date

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4-26-03

Date

PROJECT

*Antioxidant Study in HAEC cells*Notebook No. *2003-01*

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Continued From Page

*Experiment 2003-01-95*Antioxidant Testing in HAEC
Antioxidant assay using HAEC and 1000 μ M H₂O₂+DMSO

Testing dates:

No.	Test Material	6/3/03	6/4/03	6/5/03	No. Wells
		d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	
1	Hopsteiner CO2 Hop Extract (AN1082)	100	10.0	1.00	8
2	Hopsteiner Beta AromExtract Light Stable (AN1083)	100	10.0	1.00	8
3	5% Beta Hydrohop (AN1084)	100	10.0	1.00	8
4	5% Alpha Hydrohop (AN1085)	100	10.0	1.00	8
5	YC-Hop Arom (AN1086)	100	10.0	1.00	8
6	YC-Alpharich (AN1087)	100	10.0	1.00	8
7	YC-TETRA (AN1088)	100	10.0	1.00	8
8	YC-Kettle RHO (AN1089)	100	10.0	1.00	8
9	ISO-Rich (AN1090)	100	10.0	1.00	8
10	YC-Purified Alpha	100	10.0	1.00	8
total =					80

*Purpose + Procedure
203-01-30**Plate 1*

08.02.03/2 HAEC ROI

*10:100
10ul into 90ul*Antioxidant Testing in HAEC
Antioxidant assay using HAEC and 1000 μ M H₂O₂+DMSO

Testing dates:

No.	Test Material	6/3/03	6/4/03	6/5/03	No. Wells
		d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	
1	BetaTech 1% Alphahop (AN1124)	100	10.0	1.00	8
2	BetaTech 1% BetaStab 10A (AN1125)	100	10.0	1.00	8
3	BetaTech 1% Aromahop (AN1126)	100	10.0	1.00	8
4	BetaTech 1% Isohop (AN1127)	100	10.0	1.00	8
5	BetaTech 1% Redihop (AN1128)	100	10.0	1.00	8
6	BetaTech 1% Tetrahop Gold (AN1129)	100	10.0	1.00	8
7	BetaTech 1% Hexahop Gold (AN1130)	100	10.0	1.00	8
8	BetaTech 1% Hop Oil (AN1131)	100	10.0	1.00	8
9	Mg Rho 50 (RIAA) (AN1176)	100	10.0	1.00	8
10	RIAA Hop #1199 (AN1177)	100	10.0	1.00	8
total =					80

*Plate 2*Continued on Page *46*

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Signed*6-15-03*
Date

RAW

Notebook No. 2003-04

22

PROJECT 81503 lysates on Cox-1, Cox-2, iNOS, IK β Blots

Continued From Page

2003-04-22

Purpose: To look at Cox-1, Cox-2, iNOS, IK β in RAW cell lysates from 2003-03-41

Procedure:

4 Gels will be run using the Savant mini-electroblot system. Gels were removed from 4°C and rinsed in ultra-pure water. They were snapped into the savant system and 1x SDS PAGE buffer was added. See 2003-04-08 for 1x SDS PAGE buffer.

4 sets of D. Siml Eppendorf self-locking tubes were set up. Equal volume of loading buffer (Sigma S-3401 lot 21K 9279) and protein sample (volume is in experiment 2003-03-41) were added to appropriate tubes. The tubes were shut and heated @ 100°C for 4 minutes to degrade proteins using a VWR standard HeatBlock. The samples were then spun down briefly using an E.C. micromax ultra-centrifuge.

Then standard was added at 7ul to lane 1 of each gel (BioRad 161-0263 Precision plus Protein standard - unstained) lot # 96182.

Lane	2	- control
"	3	- LPS 10ug/ml
"	4	- RIAA 10ug/ml
"	5	- RIAA/LPS 10ug/10ug/ml
"	6	- Curcumin 10ug/ml
"	7	- Curcumin/RIAA 5ug/ml/5ug/ml
"	8	- Curcumin/RIAA 5ug/ml/5ug/ml
"	9	- Curcumin/RIAA/LPS 5ug/ml/5ug/ml/10ug/ml
"	10	- Caffeine 25ug/ml (used from experiment 2003-03-41)
"	11	- Caffeine/LPS 25ug/ml/10ug/ml
"	12	- Caffeine/RIAA/LPS 5ug/5ug/10ug/ml
"	13	- BOTH

Note order Change!

Continued on Page 23

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Signed *James M. Will*

9-3-03

James M. Will

9-5-03

PROJECT PE, Assay in RAW + HASCNotebook No. 2003-05
Continued From Page _____

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2003-05-42Procedure on Page 2003-05-03

C1_COXAGS_12.15.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 [50 μ M] added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
12/18/03

Compound	Comments	d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	d4 [μ g/mL]	No Wells
1. Salixalin		50	5	0.5	0.05	8
2. New Naprox		50	5	0.5	0.05	8
3. Old Naprox		50	5	0.5	0.05	8
4. Pain Out		50	5	0.5	0.05	8
5. Joint Ease		50	5	0.5	0.05	8
6. Pharma Plam		50	5	0.5	0.05	8
7. Aspirin		50	5	0.5	0.05	8
8. APHB (Cayman)		50	5	0.5	0.05	8
Total =						64

C2_COXAGS_12.16.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 [50 μ M] added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
12.16.03

Compound	Comments	d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	d4 [μ g/mL]	No Wells
1. Devia Care 600 PLUS		50	5	0.5	0.05	8
2. LYMPHODRAN ORTHOPLEX		50	5	0.5	0.05	8
3. A.R.X. ORTHOPLEX		50	5.0	0.5	0.05	8
4. ARTHRO-COMPLEX		50	5.0	0.5	0.05	8
5. ARENCAP®		50	5.0	0.5	0.05	8
6. Traumagel®		50	5.0	0.5	0.05	8
7. Indomethacin		50	5.0	0.5	0.05	8
8. Celebrex®		50	5.0	0.5	0.05	8
Total =						64

Continued on Page 43

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12-15-03
Date

[Signature]
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12-20-03
Date

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